

Inflammatory Neurodegeneration and Mechanisms of Microglial Killing of Neurons

Guy C. Brown · Jonas J. Neher

Received: 30 November 2009 / Accepted: 2 February 2010 / Published online: 2 March 2010
© Springer Science+Business Media, LLC 2010

Abstract Inflammatory neurodegeneration contributes to a wide variety of brain pathologies. A number of mechanisms by which inflammatory-activated microglia and astrocytes kill neurons have been identified in culture. These include: (1) acute activation of the phagocyte NADPH oxidase (PHOX) found in microglia, (2) expression of the inducible nitric oxide synthase (iNOS) in glia, and (3) microglial phagocytosis of neurons. Activation of PHOX (by cytokines, β -amyloid, prion protein, lipopolysaccharide, ATP, or arachidonate) causes microglial proliferation and inflammatory activation; thus, PHOX is a key regulator of inflammation. However, activation of PHOX alone causes little or no death, but when combined with iNOS expression results in apparent apoptosis via peroxynitrite production. Nitric oxide (NO) from iNOS expression also strongly synergizes with hypoxia to induce neuronal death because NO inhibits cytochrome oxidase in competition with oxygen, resulting in glutamate release and excitotoxicity. Finally, microglial phagocytosis of these stressed neurons may contribute to their loss.

Keywords Inflammation · Cell death · Nitric oxide · Peroxynitrite · Phagocytosis · iNOS · NADPH oxidase

Abbreviations

COX-2 Cyclooxygenase-2
LPS Lipopolysaccharide
LTA Lipoteichoic acid
NO Nitric oxide

NOS Nitric oxide synthase
iNOS Inducible NOS
PHOX Phagocytic NADPH oxidase
PS Phosphatidylserine
RONS Reactive oxygen and nitrogen species

Introduction

Inflammatory neurodegeneration is neuronal death/loss caused by inflammation and is a process (like energy depletion, protein aggregation, and excitotoxicity) contributing to neuronal loss or dysfunction in many different neurological diseases. In general, inflammation may have beneficial and/or detrimental effects in any particular disease and in any particular phase of a disease. The beneficial effects are mainly due to elimination of pathogens, clearing debris, and aiding repair, and the detrimental effects are probably unintended side effects of the beneficial processes. There is evidence that brain inflammation may contribute to the pathology of both chronic neurodegenerative diseases (Alzheimer's, Parkinson's, multiple sclerosis, and AIDS dementia) as well as acute pathologies such as stroke, brain trauma, and meningitis [1–6]. These pathologies have different causes and consequences, but they all involve brain inflammation, and there is evidence that blocking inflammation can either delay onset or reduce symptoms [1–6]. However, there are different types or modes of inflammation, and it is important to understand why inflammation is sometimes damaging and at other times protective so that interventions can be designed to prevent one but not the other.

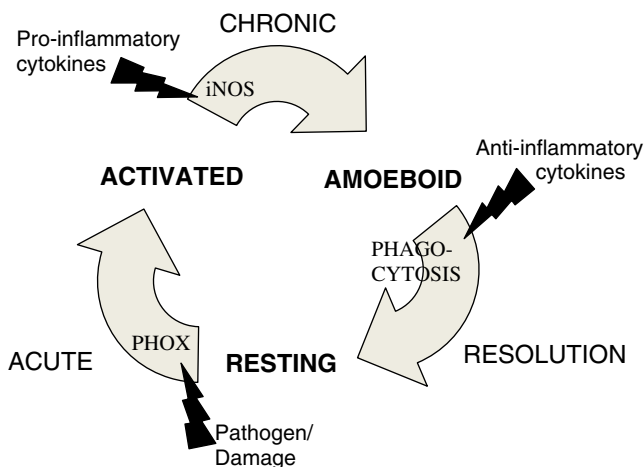
Inflammation can damage the brain in a variety of ways, including: (1) inflammation in the vascular wall may drive

G. C. Brown (✉) · J. J. Neher
Department of Biochemistry, University of Cambridge,
Tennis Court Road,
Cambridge CB2 1QW, UK
e-mail: gcb@mole.bio.cam.ac.uk

atherosclerosis, leading to stroke and vascular dementia; (2) inflammation in the blood–brain barrier may compromise barrier function and allow leukocytes and antibodies into the brain; (3) antibodies generated against brain antigens may induce immune attack as occurs in multiple sclerosis; (4) inflammation may induce brain edema (swelling); (5) some types of inflammation may suppress neurogenesis; and (6) pathogens, protein aggregates, or damaged neurons may inflammatory activate glia, which may then kill neurons. It is this last type of damage, common to many brain pathologies, that we shall be concerned with here.

Microglia, the brain's resident macrophages, are the predominant immune cells in the healthy brain. This is due to the brain being shielded by the blood–brain barrier (BBB) which limits the entry of peripheral cells and substances. Brain inflammation, which occurs behind the BBB, therefore differs from inflammation in the periphery by the relative absence of leukocytes (including neutrophils, monocytes, B cells, and T cells) and antibodies. However, it is now recognized that there is a limited traffic of these across the barrier, and this traffic can be increased by inflammation which can recruit leukocytes into the brain [7]. Nevertheless, microglia are key cells in brain inflammation and inflammatory neurodegeneration [5, 8].

Brain inflammation may be divided into three phases (acute, chronic, and resolution) in which the microglia are largely in three different morphologies (resting, activated, and amoeboid/phagocytic, see Scheme 1). The healthy,

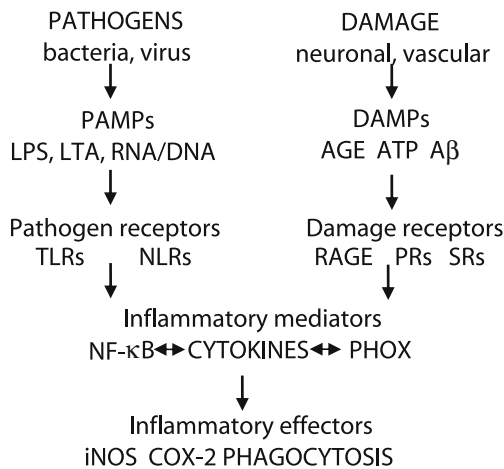


Scheme 1 Hypothetical relation of the three phases of inflammation to the three states of microglial activation, driven by three types of signal, acting through three effectors. Microglial PHOX activation by pathogens or damage drives acute activation, which results in pro-inflammatory cytokine production and iNOS expression. Chronic activation may result in amoeboid, highly phagocytic microglia and a switch to anti-inflammatory cytokines driving the resolution phase. Different phases may coexist in the brain, and PHOX activation may occur in the chronic phase, but (in the presence of iNOS) will then result in peroxynitrite production (rather than superoxide and hydrogen peroxide)

non-inflamed brain contains almost entirely “resting” microglia which are highly ramified, with a small, static cell body, but with dynamic and branched processes actively seeking out (1) pathogens and (2) damage in the brain [9]. Pathogen receptors (pattern recognition receptors) include the Toll-like receptors (TLRs) and the NOD-like receptors (NLRs), and generally recognize cell wall components or RNA/DNA of pathogens (Scheme 2). Damage (or “danger”) receptors include scavenger receptors, purinergic receptors, receptor for advanced glycation endproducts (RAGE), and TLRs, and these recognize components released from stressed or damaged host cells, including ATP, aggregated β -amyloid, and heat shock proteins [5]. Engagement of these receptors induces signal cascades that will (after several hours) produce the “chronically activated” state of microglia due to expression of new proteins, including iNOS, COX-II, and MHC-II, the latter enabling microglia to present antigens to T cells. Activation is accompanied by partial rounding up and mobility of the cells, proliferation, and the expression and release of pro-inflammatory cytokines, including TNF α , IL-1 β , and IL-6. These cytokines activate other microglia and astrocytes. Most of the expression changes are a result of activation of the transcription factor NF- κ B via phosphorylation-induced activation of I κ B kinase.

Prior to the induction of gene expression, the microglia are in the “acute” phase of activation triggered by receptor ligation. Receptor stimulation induces PKC activity, causing rapid activation of the phagocyte NADPH oxidase (PHOX) which in turn contributes to NF- κ B activation. After induced gene expression, the microglia are in a chronic state of activation, which may be maintained by the pro-inflammatory cytokine release and the continued presence of pathogen/damage. However, the chronic state of activation may progress to a “resolution phase” where microglia are amoeboid, highly phagocytic, and produce anti-inflammatory cytokines (including IL-10 and TGF β) in order to resolve the inflammation and clear up the mess. This three-phase division (acute, chronic, and resolution) of microglial inflammation is simplistic as there are alternative modes of activation and alternative ways of classifying microglial activation [8]. However, the progression through these three inflammatory states may be a mechanism common to microglial activation.

Activated microglia can kill and/or remove pathogens, but they may also kill neurons. The mechanisms by which they kill neurons are complex and not fully understood [5]. We will restrict ourselves to reviewing the roles of three different processes (NADPH oxidase activation, iNOS expression, and phagocytosis) corresponding to the three phases of inflammation (acute, chronic, and resolution phases), with particular emphasis on our own research in this area.



Scheme 2 How inflammation is induced by pathogens or damage. Pathogens are recognized by pathogen receptors, mainly Toll-like receptors (*TLRs*) or NOD-like receptors (*NLRs*), on host cells which recognize pathogen-associated molecular patterns (*PAMPs*) on the pathogen, such as bacterial cell wall components lipopolysaccharide (*LPS*) or lipoteichoic acid (*LTA*), or bacterial/viral RNA or DNA. Damage produces damage-associated molecular patterns (*DAMPs*) such as advanced glycation endproducts (*AGE*) that are recognized by the receptor for *AGE* (*RAGE*) or extracellular nucleotides such as *ATP* that are recognized by certain purinergic receptors (*PRs*) or aggregated proteins such as beta-amyloid (*Aβ*) that are recognized by *RAGE* and scavenger receptors (*SRs*). Engagement of receptors results in the activation of inflammatory pathways such as the phagocyte NADPH oxidase (*PHOX*) and *NF-κB*, and causes cytokine and reactive oxygen production and release, which feeds back to cause further and spreading inflammation. This in turn causes expression of inflammatory effectors: inducible NO synthase (*iNOS*), cyclooxygenase-2 (*COX-2*), as well as phagocytosis

The Phagocyte NADPH Oxidase

Phagocytic cells such as neutrophils, macrophages, and microglia have a specific NOX (NADPH oxidase) known as *PHOX* (phagocytic oxidase), consisting of subunits gp91, p22, p47, p67, p40, and Rac. In the healthy, non-inflamed brain, *PHOX* is expressed at high levels in microglia (and possibly at low levels in astrocytes and neurons). However, *PHOX* is not active unless acutely stimulated by, for example, *TNF-α*, *IL-1β*, chemokines, arachidonate, *β*-amyloid, lipopolysaccharide (*LPS*), *ATP*, or phagocytosis. When activated, it rapidly produces high levels of superoxide extracellularly, which may either dismutate to hydrogen peroxide (catalyzed by extracellular superoxide dismutase) or react with NO to produce peroxynitrite [10]. These oxidants contribute to the killing of pathogens by phagocytes, but may also damage neurons.

Importantly, however, activation of *PHOX* alone causes no apparent neuronal death [11], but stimulates the microglia to proliferate [12], produce *TNFα* and *IL-1β* [13, 14], and express *iNOS* [14]. For example, we showed that

fibrillar *β*-amyloid induces microglia to proliferate and produce cytokines via the activation of *PHOX* [13]. In general, *H₂O₂* from *PHOX* leads to cell proliferation and activation by oxidation of the active site cysteine residue of protein tyrosine phosphatases, causing inhibition of these phosphatases and thus amplification of mitotic and pro-inflammatory tyrosine kinase cascades [15]. *H₂O₂* may also activate matrix metalloproteinases which can amplify inflammation. *PHOX* has been shown by several labs (particularly Block et al. [5]) to be a key regulator of inflammatory activation of microglia. Correspondingly, inhibition of *PHOX* is sufficient to prevent activation of microglia and thus is a target for anti-inflammatory strategies.

Inducible NO Synthase

Inducible NO synthase (*iNOS*) is not normally expressed in the brain, but inflammatory mediators such as *LPS* and cytokines cause its expression in microglia and astrocytes [16] and possibly in neurons [17]. Once expressed, *iNOS* produces high levels of NO continuously (up to 1 *μM* NO from microglia or astrocytes [10, 18]). We showed that these high levels of NO induce neuronal death by causing inhibition of mitochondrial cytochrome oxidase in neurons [18, 19]. NO inhibition of neuronal respiration caused neuronal depolarization and glutamate release, followed by excitotoxicity via the NMDA receptor [18, 20–23]. This excitotoxicity may be potentiated by a second mechanism as NO from *iNOS* results in glutamate release from astrocytes via calcium release from intracellular stores stimulating exocytosis of vesicular glutamate [24]. Thus, inflammatory-activated astrocytes maintained a relatively high extracellular glutamate level [24]. This level of glutamate may be insufficient to induce excitotoxicity alone, but it has been shown that non-toxic levels of extracellular glutamate can become toxic to neurons if in addition the mitochondrial respiratory chain is inhibited, possibly because such inhibition may partially depolarize the neurons, activating the NMDA receptor if glutamate is present outside [25]. However, high glutamate may impose an increased metabolic demand on neurons so that if metabolism is inhibited, this may suffice to push the cell over some toxic threshold. Such mechanisms may contribute to how NO kills neurons.

However, the above mechanism requires relatively high levels of NO or *iNOS* expression, as occurs in the presence of *IFNγ* which dramatically potentiates *iNOS* expression [26] and changes microglial phenotype in multiple ways. In contrast, *iNOS* can be expressed at low levels in vitro [11] or in vivo [27] apparently with little or no neuronal death. Indeed, NO from *iNOS* may be protective by blocking

brain cell death [28, 29]. On the other hand, low levels of iNOS expression may synergise with other conditions to induce cell death [30]. For example, hypoxia strongly synergises with NO or iNOS expression to induce neuronal death via respiratory inhibition [31]. This is because NO is a competitive inhibitor of cytochrome oxidase, the NO competing with oxygen for binding to cytochrome oxidase [19, 31] so that NO greatly increases the apparent K_M of neuronal respiration for oxygen. This sensitisation to hypoxia is potentially important in stroke, trauma, vascular dementia, Alzheimer's, and brain aging where both inflammation and hypoxia may coexist (Scheme 3). Note, however, that O_2 is a substrate for all NO synthases with apparent K_M of 4, 130, and 350 μM O_2 for eNOS, iNOS, and nNOS, respectively, [32] so that nNOS and iNOS may be substrate-limited by oxygen. On the other hand, O_2 also promotes NO consumption by NO reaction with oxygen, superoxide, oxyhemoglobin, or oxymyoglobin [33] so that the dependence of NO level on O_2 is complex and poorly understood [34].

iNOS and PHOX

As described above, neither activation of NADPH oxidase (NOX/PHOX) nor production of (moderate levels of) nitric oxide through iNOS expression is sufficient to induce neurotoxicity. However, we found that when both were activated together, this resulted in apparent apoptosis of most neurons mediated by peroxynitrite (Scheme 3). We showed that inflammatory neurodegeneration induced by $TNF\alpha$, $IL-1\beta$, prion peptide, LPS, $IFN\gamma$, arachidonate, ATP, and/or PMA was mediated by this dual-key (iNOS and PHOX) mechanism of inflammatory neurodegeneration in particular conditions [11]. Simultaneous activation of

PHOX and iNOS in microglia resulted in the disappearance of NO [10], appearance of peroxynitrite [10, 11], and apoptosis of co-cultured neurons that was prevented by inhibitors of iNOS or PHOX or by scavengers of superoxide or peroxynitrite [11].

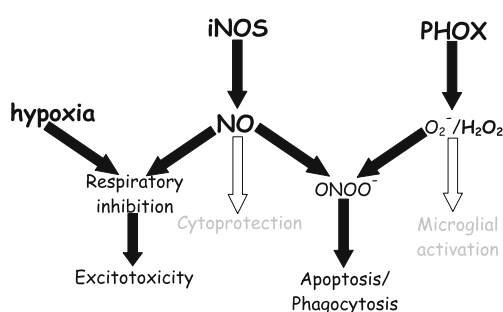
Phagocytosis

Lipoteichoic acid (LTA) is a cell wall component of Gram-positive bacteria which may trigger inflammatory neurodegeneration in bacterial meningitis. We showed that LTA induces iNOS in glia via Toll-like receptor 2 [26] and results in loss of neurons in glial-neuronal co-cultures. This neuronal loss is prevented by scavengers of superoxide or peroxynitrite [35]. However, intriguingly, there is no measurable neuronal death in these cultures, only neuronal loss presumably due to phagocytosis [35].

How apoptosis causes cells to die is, surprisingly, not always clear, but in vivo, the main means is by triggering exposure of "eat me" signals (such as phosphatidylserine) on the cell surface which induce phagocytes to eat the cell. Reactive oxygen and nitrogen species (RONS) can induce PS exposure directly via activation of the phospholipid scramblase and inactivation of amino phospholipid translocase (flippase) [36] or indirectly via induction of apoptosis [37, 38]. However, it is important to realize that phosphatidylserine exposure can be reversible [39] including NO-induced phosphatidylserine exposure of neurons [40]. Furthermore, RONS-induced PS exposure by neurons may induce activation of microglia [37, 38], which may then phagocytose the neurons. Thus, it is possible that microglial phagocytosis may actively contribute to neuronal death.

Conclusions

We have discussed the nature of inflammatory neurodegeneration and outlined the contributions of PHOX, iNOS, and phagocytosis to microglial killing of neurons during inflammation. A variety of other neurotoxic factors released from activated microglia have been identified, including glutamate, $TNF\alpha$, Fas ligand, cathepsin B, and other proteases. Much future work is required to elucidate the contribution of all these processes to particular brain diseases. This would be greatly aided by drugs that specifically inhibit these targets within the brain without toxicity. We also need to know the relative contributions of inflammation, energy depletion, protein aggregation, and excitotoxicity to the pathology of different brain diseases and to assess the extent to which blocking these processes individually or in combination blocks pathology.



Scheme 3 Mechanisms of inflammatory neurodegeneration. Hypoxia, iNOS expression, or PHOX activation may be relatively benign, or even cytoprotective, when present alone. However, hypoxia combined with NO results in neuronal death via respiratory inhibition, and PHOX activation combined with iNOS expression results in peroxynitrite production, inducing neuronal apoptosis at high levels, but loss of neurons by microglial phagocytosis at lower levels

Acknowledgments Relevant research in our laboratory has been funded by the Wellcome Trust, Medical Research Council, Alzheimer's Research Trust, and European Union.

References

- Klegeris A, McGeer EG, McGeer PL (2007) Therapeutic approaches to inflammation in neurodegenerative disease. *Curr Opin Neurol* 20:351–357
- Zipp F, Aktas O (2006) The brain as a target of inflammation: common pathways link inflammatory and neurodegenerative diseases. *Trends Neurosci* 29:518–527
- Lucas SM, Rothwell NJ, Gibson RM (2006) The role of inflammation in CNS injury and disease. *Br J Pharmacol* 147 (Suppl 1):S232–S240
- Brown GC, Bal-Price A (2003) Inflammatory neurodegeneration mediated by nitric oxide, glutamate, and mitochondria. *Mol Neurobiol* 27:325–355
- Block ML, Zecca L, Hong JS (2007) Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nat Rev Neurosci* 8:57–69
- Wyss-Coray T (2006) Inflammation in Alzheimer disease: driving force, bystander or beneficial response? *Nat Med* 12:1005–1015
- Engelhardt B, Ransohoff RM (2005) The ins and outs of T-lymphocyte trafficking to the CNS: anatomical sites and molecular mechanisms. *Trends Immunol* 26:485–495
- Ransohoff RM, Perry VH (2009) Microglial physiology: unique stimuli, specialized responses. *Annu Rev Immunol* 27:119–145
- Hanisch UK, Kettenmann H (2007) Microglia: active sensor and versatile effector cells in the normal and pathologic brain. *Nat Neurosci* 10:1387–1394
- Bal-Price A, Brown GC (2002) Stimulation of the NADPH oxidase in activated rat microglia removes nitric oxide but induces peroxynitrite production. *J Neurochem* 80:73–80
- Mander P, Brown GC (2005) Activation of microglial NADPH oxidase is synergistic with glial iNOS expression in inducing neuronal death: a dual-key mechanism of inflammatory neurodegeneration. *J Neuroinflamm* 2:20
- Mander PK, Jekabsone A, Brown GC (2006) Microglia proliferation is regulated by hydrogen peroxide from NADPH oxidase. *J Immunol* 176:1046–1052
- Jekabsone A, Mander PK, Tickler A, Sharpe M, Brown GC (2006) Fibrillar beta-amyloid peptide A β 1–40 activates microglial proliferation via stimulating TNF- α release and H₂O₂ derived from NADPH oxidase: a cell culture study. *J Neuroinflamm* 3:24
- Pawate S, Shen Q, Fan F, Bhat NR (2004) Redox regulation of glial inflammatory response to lipopolysaccharide and interferon- γ . *J Neurosci Res* 77:540–551
- Chan EC, Jiang F, Peshavariya HM, Dusting GJ (2009) Regulation of cell proliferation by NADPH oxidase-mediated signaling: potential roles in tissue repair, regenerative medicine and tissue engineering. *Pharmacol Ther* 122:97–108
- Murphy S (2000) Production of nitric oxide by glial cells: regulation and potential roles in the CNS. *Glia* 29:1–13
- Heneka MT, Feinstein DL (2001) Expression and function of inducible nitric oxide synthase in neurons. *J Neuroimmunol* 114:8–18
- Bal-Price A, Brown GC (2001) Inflammatory neurodegeneration mediated by nitric oxide from activated glia, inhibiting neuronal respiration, causing glutamate release and excitotoxicity. *J Neurosci* 21:6480–6491
- Brown GC, Cooper CE (1994) Nanomolar concentrations of nitric oxide reversibly inhibit synaptosomal respiration by competing with oxygen at cytochrome oxidase. *FEBS Lett* 356:295–298
- McNaught K, St P, Brown GC (1998) Nitric oxide causes glutamate release from brain synaptosomes following inhibition of mitochondrial function. *J Neurochem* 70:1541–1546
- Stewart VC, Heslegrave AJ, Brown GC, Clark JB, Heales SJ (2002) Nitric oxide-dependent damage to neuronal mitochondria involves the NMDA receptor. *Eur J Neurosci* 15:458–464
- Golde S, Chandran S, Brown GC, Compston A (2002) Different pathways for iNOS-mediated toxicity in vitro dependent on neuronal maturation and NMDA receptor expression. *J Neurochem* 82:269–282
- Jekabsone A, Neher J, Borutaite V, Brown GC (2007) Nitric oxide from neuronal nitric oxide synthase sensitises neurons to hypoxia-induced death via competitive inhibition of cytochrome oxidase. *J Neurochem* 103:346–356
- Bal-Price A, Moneer Z, Brown GC (2002) Nitric oxide induces rapid, calcium-dependent release of vesicular glutamate and ATP from cultured rat astrocytes. *Glia* 40:312–323
- Novelli A, Reilly JA, Lysko PG, Henneberry RC (1988) Glutamate becomes neurotoxic via the *N*-methyl-D-aspartate receptor when intracellular energy levels are reduced. *Brain Res* 451:205–212
- Kinsner A, Boveri M, Hareng L, Traub S, Brown GC, Coecke S, Hartung T, Bal-Price A (2006) Highly purified lipoteichoic acid induced proinflammatory signalling in primary culture of rat microglia through Toll-like receptor 2: selective potentiation of nitric oxide production by muramyl dipeptide. *J Neurochem* 99:596–607
- Han HS, Qiao Y, Karabiyikoglu M, Giffard RG, Yenari MA (2002) Influence of mild hypothermia on inducible nitric oxide synthase expression and reactive nitrogen production in experimental stroke and inflammation. *J Neurosci* 22:3921–3928
- Takuma K, Phuaphong P, Lee E, Mori K, Baba A, Matsuda T (2001) Anti-apoptotic effect of cGMP in cultured astrocytes: inhibition by cGMP-dependent protein kinase of mitochondrial permeable transition pore. *J Biol Chem* 276:48093–48099
- Cho S, Park EM, Zhou P, Frys K, Ross ME, Iadecola C (2005) Obligatory role of inducible nitric oxide synthase in ischemic preconditioning. *J Cereb Blood Flow Metab* 25:493–501
- Borutaite V, Brown G (2005) What else has to happen for nitric oxide to induce cell death? *Biochem Soc Trans* 33:1394–1396
- Mander P, Borutaite V, Moncada S, Brown GC (2005) Nitric oxide from glial iNOS sensitizes neurons to hypoxic death via mitochondrial respiratory inhibition. *J Neurosci Res* 79:208–215
- Santolini J, Meade AL, Stuehr DJ (2001) Differences in three kinetic parameters underpin the unique catalytic profiles of nitric oxide synthases I, II, and III. *J Biol Chem* 276:48887–48898
- Liu X, Miller MJ, Joshi MS, Thomas DD, Lancaster JR Jr (1998) *Proc Natl Acad Sci USA* 95:2175–2179
- Brown GC, Borutaite V (2006) Interactions between nitric oxide, oxygen, reactive oxygen species and reactive nitrogen species. *Biochem Soc Trans* 34:953–956
- Kinsner A, Pilotto V, Deininger S, Brown GC, Coecke S, Hartung T, Bal-Price A (2005) Inflammatory neurodegeneration induced by lipoteichoic acid from *Staphylococcus aureus* is mediated by glia activation, nitrosative and oxidative stress, and caspase activation. *J Neurochem* 95:1132–1143
- Tyurina YY, Basova LV, Konduru NV, Tyurin VA, Potapovich AI, Cai P, Bayir H, Stoyanovsky D, Pitt BR, Shvedova AA, Fadeel B, Kagan VE (2007) Nitrosative stress inhibits the aminophospholi-

- pid translocase resulting in phosphatidylserine externalization and macrophage engulfment: implications for the resolution of inflammation. *J Biol Chem* 282:8498–8509
37. Kang JQ, Chong ZZ, Maiese K (2003) Critical role for Akt1 in the modulation of apoptotic phosphatidylserine exposure and microglial activation. *Mol Pharmacol* 64:557–569
38. Kang JQ, Chong ZZ, Maiese K (2003) Akt1 protects against inflammatory microglial activation through maintenance of membrane asymmetry and modulation of cysteine protease activity. *J Neurosci Res* 74:37–51
39. Balasubramanian K, Mirnikjoo B, Schroit AJ (2007) Regulated externalization of phosphatidylserine at the cell surface: implications for apoptosis. *J Biol Chem* 282:18357–18364
40. Maiese K, Vincent AM (2000) Membrane asymmetry and DNA degradation: functionally distinct determinants of neuronal programmed cell death. *J Neurosci Res* 59:568–580